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(54) Title: RECOMBINANT HUMAN ERYTHROPOIETIN WITH ADVANTAGEOUS GLYCOSYLATION PROFILE (57) Abstract <p>Described are polypeptides having part or all of the primary structural conformation and biological property of erythropoietin and having an improved in vivo half-life and biological activity due to a modified glycosylation profile. Also provided are DNA sequences encoding the amino acid sequence of said polypeptide operatively linked to regulatory elements which allow for the expression of said DNA sequence in the eukaryotic host cell as well as vectors comprising such DNA sequences. Furthermore, host cells comprising the aforementioned DNA sequences and vectors and their use for the production of the aforescribed polypeptide are described. Pharmaceutical and diagnostic compositions are provided comprising the polypeptides, DNA sequences or vectors of the invention. Described is also the use of the aforescribed polypeptides, DNA sequences and vectors for the preparation of pharmaceutical compositions for treating all kinds of anaemia caused by a lack of erythropoietin.</p>		

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RECOMBINANT HUMAN ERYTHROPOIETIN WITH ADVANTAGEOUS GLYCOSYLATION PROFILE

The present invention relates to polypeptides having part or all of the primary structural conformation of erythropoietin and having an improved in vivo half-life and biological activity due to a modified glycosylation profile. The present invention also provides DNA sequences encoding the amino acid sequence of said polypeptides operatively linked to regulatory elements which allow for the expression of said DNA sequence in eukaryotic host cells as well as vectors comprising such DNA sequences. The present invention also relates to host cells comprising the aforementioned DNA sequences and vectors and their use for the production of the aforescribed polypeptides.

Furthermore, the present invention relates to pharmaceutical and diagnostic compositions comprising the aforementioned polypeptides, DNA sequences and vectors. The present invention also relates to the use of the aforescribed polypeptides, DNA sequences and vectors for the preparation of pharmaceutical compositions for treating all kinds of anaemia caused by a lack of erythropoietin.

The erythrocyte is by far the most common type of cell in the blood. When mature, it is packed full of hemoglobin and contains practically none of the usual cell organelles. In an erythrocyte of an adult mammal, even the nucleus, endoplasmic reticulum, mitochondria, and ribosomes are absent, having been extruded from the cell in the course of its development. The erythrocyte, therefore, cannot grow or divide; the only possible way of making more erythrocytes is by means of stem cells. Furthermore, erythrocytes have a limited life span of about 120 days in humans. Worn-out erythrocytes are phagocytosed and digested by macrophages in the liver and spleen, which remove more than 10^{11} senescent erythrocytes in every human being per day. A lack of oxygen or a shortage of erythrocytes stimulates cells in the kidney to synthesize

and secrete increased amounts of erythropoietin into the blood-stream. The erythropoietin in turn stimulates the production of more erythrocytes. Since the change in the rate of release of new erythrocytes into the blood-stream is observed as early as 1 or 2 days after an increase in erythropoietin levels in the blood-stream, the hormone must act on cells that are very close precursors of the mature erythrocytes. The cells that respond to erythropoietin can be identified by culturing bone marrow cells in a semisolid matrix in the presence of erythropoietin. In a few days colonies of about 60 erythrocytes appear, each founded by a single committed erythrocyte progenitor cell. This cell is known as an erythrocyte colony-forming cell, or CFC-E, and gives rise to mature erythrocytes after about six division-cycles or less. The CFC-Es do not yet contain hemoglobin, and they are derived from an earlier type of progenitor cell whose proliferation does not depend on erythropoietin. CFC-Es themselves depend on erythropoietin for their survival as well as for proliferation: if erythropoietin is removed from the cultures, the cells rapidly undergo programmed cell death. Erythropoietin as other colony stimulating factor is a glycoprotein that acts at low concentration (about 10^{-12} M) by binding to specific cell-surface receptors. These receptors belong to a large receptor family, the so-called "cytokine receptor family", whose members are usually composed of two or more subunits, one of which is frequently shared among several receptor types. Mature human erythropoietin is a glycoprotein with a molecular weight of 34 to 38 kD and consists of 166 amino acids (AS) and the glycosyl residue accounts for about 40 % of the molecular weight. Since erythropoietin is required for the renewal of erythrocytes, this hormone is essential for the quality of life, especially of patients, which suffer from anaemia and hypoxia, due to reduced numbers of red blood cells which can be caused by, e.g., dialysis or through reduction of erythroid precursor cells as a consequence of therapies based on the suppression of cellular proliferation or by inborne or aquired insufficiency of erythropoietin production. The identification of the human gene encoding erythropoietin made it possible to recombinantly express this protein in heterologous host cells and to provide sufficient amounts of recombinant human erythropoietin (rhuEpo) for the treatment of the diseases mentioned.

However, apart from the primary structure of the protein the structure of the sugar side chains of the molecule is of particular importance for the interaction of Epo within the organism. For example, desialylated Epo shows no effect upon application in animals. It nevertheless binds to the receptor and stimulates precursor cells. The activity decrease of asialo-Epo in vivo can be explained by the fact that it is removed in the liver via receptors with a specificity for galactosyl residues which are susceptible in desialylated Epo.

The wildtype Epo, which has been used therapeutically, has in some patients the effect of increasing the blood pressure, which is disadvantageous in therapy. It is to be assumed that Epo also is integrated in the blood pressure regulation. Therefore, it is desirable to have proteins with the physiological effect of Epo at one's disposal which do, however, not show these undesired properties but which nevertheless stimulate the differentiation and division rate of precursor cells to erythrocytes. A further side effect of Epo found in some patients is the stimulation of the megakaryocytes for the formation of thrombocytes. Therefore, there is potential danger of thrombosis during the therapy with Epo, which then has to be discontinued immediately. In this case, a higher specificity of the Epo used would be desirable.

Thus, the technical problem underlying the present invention is to provide rhuEpo having an improved biological activity and in-vivo half-life compared to naturally occurring and rhuEpo so far available.

The solution to the above technical problem is achieved by providing the embodiments characterized in the claims.

Accordingly, the invention relates to a polypeptide having part or all of the primary structural conformation of erythropoietin possessing the biological property of causing bone marrow cells to increase production of reticulocytes and red blood cells and to increase haemoglobin synthesis or iron uptake, said polypeptide being the product of

eukaryotic expression of an exogenous DNA sequence and having the following physiochemical properties:

- (i) the amino acid sequence comprises the amino acid sequence given in SEQ ID No. 1 or any fragment or derivative thereof by way of amino acid deletion, substitution, insertion, addition and/or replacement of the amino acid sequence given in SEQ ID No. 1, wherein at least one of the consensus N-linked glycosylation sites is modified to other than a consensus N-linked glycosylation site;
- (ii) it is glycosylated;
- (iii) greater 5% of the N-glycan structures are sulfated; and
- (iv) the ratio Z^* of the total N-glycan charge Z to the number of N-glycosylation sites is greater than 170.

Human Epo has one O-linked (at Ser126 of SEQ ID NO: 1) and three N-linked glycosyl residues (at Asn 24, Asn 38 and Asn 83 of SEQ ID NO: 1), which are sialylated¹. It has been demonstrated that proper sialylation of these sugar residues is important for the in vivo half-life and, subsequently, for the efficacy of Epo². Furthermore, it has been demonstrated that different glycosylation sites of rhuEpo are glycosylated differently concerning the complexity of the sugar structures and sialylation^{7,8}. The analysis of genetically engineered glycomutins showed, that elimination of glycosylation site Asn24 of the rhuEpo molecule (SEQ ID NO.: 1) resulting in the glycomutins rhuEpo (Gln24) (SEQ ID NO.: 2) is advantageous for both expression and in vivo efficacy⁴. It has now been surprisingly found that rhuEpos that have a modified glycosylation profile in terms of sulfated N-glycan structures and N-glycan charge number per glycosylation site have an improved in-vivo half-life and biological activity compared to rhuEpo or any other rhuEpo glycomutins so far described^{3,4}. The present invention is based on the finding that the glycosylation profile of the mutins rhuEpo (Gln24) (SEQ ID NO.: 2) can even be improved by expression of the corresponding DNA sequence under the control of the regulatory elements of the Bovine Papillomavirus 1 (BPV-1) vector pHOEBE 40-7

in CHO cells. Although lacking one N-glycosylation site, the polypeptide (SEQ ID NO: 2) comprises a total charge number higher than rhuEpo(wt), and even higher than previously described rhuEpo(Gln24)^{4,5,8}, resulting in a significantly higher Z*; see Table II of Example 9. The polypeptide of the invention has a high amount of sialylated/sulfated sugar structures per glycosylation site, reducing the clearance of the molecule by the liver-specific asialogalactosyl-receptors. Furthermore, the same biological effect of a certain dose of rhuEpo(wt) is obtained with a lower dose of the polypeptide of the invention; see Fig.1 B. Additionally, as human urinary Epo was found to contain significant amounts of sulfated oligosaccharides⁹ the polypeptide of the invention resembles more closely the human urinary Epo than any of the other rhuEpo so far available.

The potential exists, in the use of recombinant DNA technology, for the preparation of various derivatives of the polypeptide of the invention, variously modified by resultant single and/or multiple amino acid deletion(s), substitution(s), insertion(s), addition(s) and/or replacement(s), for example by means of site directed mutagenesis of the underlying DNA. Recombinant DNA technology is well known to those skilled in the art. Included is the preparation of derivatives retaining the primary structural confirmation of erythropoietin possessing the biological property of causing bone marrow cells to increase production of erythrocytes and red blood cells and to increase hemoglobin synthesis or iron uptake retaining the essential glycosylation profile, namely that greater than 5 % of the N-glycan structures are sulfated and the ratio Z* of the total N-glycan charge Z to the number of N-glycosylation sites is greater than 170.

In a preferred embodiment the invention relates to the afore-mentioned polypeptide, wherein at least one of the consensus N-glycosylation sites is deleted and/or is replaced with (a) different amino acid(s).

In another preferred embodiment, at least one of the consensus N-glycosylation sites is added to the polypeptide as described above.

In a particularly preferred embodiment, the glycosylation site at the amino acid position 24 (Asn) of the amino acid sequence given in SEQ ID NO: 1 is deleted, preferably by replacing the amino acid Asn at position 24 of the amino acid sequence shown in SEQ ID NO: 1 with the amino acid Gln shown for example in the amino acid sequence of SEQ ID NO: 2.

As described above, the polypeptide of the invention has a ratio Z^* of the total N-glycane charge Z to the number of N-glycosylation sites greater than 170. In a preferred embodiment Z^* is greater than 180, e.g. 183, preferably greater than 190, e.g. 194.

Furthermore, the present invention relates to a DNA sequence encoding the amino acid sequence of the aforescribed polypeptides operatively linked to regulatory elements of Bovine Papillomavirus 1 (BPV-1) which allow for the expression of said DNA in a eukaryotic host cell. Experiments which had been carried out in the scope of the present invention revealed that when the mutein rhuEpo(Gln24) is expressed in CHO cells via a BPV-1 expression system the glycosylation pattern is significantly different from mutein rhuEpo(Gln24) expressed in CHO cells via the pABE-40-7 expression vector; see Table II, C3 and C4/C5. Using the BPV-1 expression system, rhuEpo(Gln24) comprises higher sialylation and a new quality of chargement through sulfatation; see Table II. Besides the expected recombinant protein, BPV-1 expression vectors are able to express several further functional activities, e.g. transacting factors and others, that can modulate the cellular activities of the host system. It is surprising, however, that it obviously can also modulate the pattern of posttranslational modifications, e.g. the glycosylation or sulfatation of rhuEpo(Gln24) in CHO cells.

In a preferred embodiment, the aforementioned DNA sequence further comprises regulatory elements of the metallothioneine 1 (MT-1) gene. Further examples of possible regulatory elements are viral regulatory elements such as SV40 promoter and enhancer elements.

The present invention also relates to vectors, preferably expression vectors, comprising a DNA sequence as described above.

In a preferred embodiment, the expression vector is BPV-1 vector pPHOEBE-40-7 described in the examples hereinafter.

The present invention further relates to host cells comprising a DNA sequence or a vector of the invention. The DNA sequence or vector of the invention which is present in the host cell may either be integrated into the genome of the host cell or may be maintained in some form extrachromosomally. The host cell can be any eukaryotic cell, such as CHO, BHK, C127i and others. Preferred host cells are CHO cells.

Another subject of the invention is a method for the production of the polypeptide of the invention having part or all of the primary structural conformation of erythropoietin possessing the biological property of causing bone marrow cells to increase production of reticulocytes and red blood cells and to increase hemoglobin synthesis or iron uptake, said method comprising culturing a host cell of the invention, and optionally isolating said polypeptide from the culture.

Depending on the specific constructs and conditions used, the polypeptide may be recovered from the cells, from the culture medium or from both. For a person skilled in the art it is well known that it is not only possible to express a native polypeptide but also to express the polypeptide as a fusion protein or to add signal sequences directing the polypeptide to specific compartments of the host cell, ensuring secretion of the polypeptide into the culture medium etc. Preferably, the polypeptide of the invention is

purified by affinity chromatography, using a monoclonal antibody specific for the huEpo-receptor binding site on the rhuEpo molecule¹⁰. Such monoclonal antibodies can be obtained according to conventional methods known in the art.

Thus, the present invention also relates to the polypeptide obtainable by the aforementioned method. The polypeptide of the invention is characterized by its increased half-life and bioactivity in-vivo compared to the same polypeptide having part or all of the primary structural confirmation of erythropoietin possessing the biological property of bone marrow cells to increase the production of reticulocytes and red blood cells and to increase hemoglobin synthesis or iron uptake but which does not have the advantageous glycosylation profile as described above for the polypeptide of the present invention.

Moreover, the present invention relates to a pharmaceutical composition comprising at least one of the afore-mentioned polypeptides, DNA sequences and/or vectors of the invention either alone or in combination, and optionally a pharmaceutically acceptable carrier or excipient. Examples of suitable pharmaceutical carriers are well known in the art and include phosphate buffered saline solutions, water, emulsions, such as oil/water emulsions, various types of wetting agents, sterile solutions etc. Compositions comprising such carriers can be formulated by conventional methods. The pharmaceutical compositions can be administered to the subject at a suitable dose. The dosage regimen will be determined by the attending physician considering the condition of the patient, the severity of the disease and other clinical factors. Suitable doses range, for example, from 1,000 to 10,000 units, preferably 3,000 to 6,000 units and are more preferably 4,000 units. Progress can be monitored by periodic assessment of hematocrit, number of reticulocytes, number of red blood cells, and the patient's general state of health. Administration of the suitable compositions may be effected by different ways, e.g. by intravenous, intraperitoneal, subcutaneous, intramuscular, topical or intradermal administration.

The invention also relates to a diagnostic composition comprising at least one of the afore-mentioned polypeptides, DNA sequences and/or vectors either alone or in combination, and optionally suitable means for detection. Said diagnostic composition may be used for methods for detecting anti-human Epo antibodies or Epo receptors. The polypeptide of the invention comprised in a diagnostic composition may be coupled to any reporter system such as peroxidase or ^{99}Tc .

In a further embodiment the invention relates to the use of at least one of the afore-mentioned polypeptides, DNA sequences and/or vectors either alone or in combination for the preparation of a pharmaceutical composition for treating all kinds of anaemia caused by a lack of erythropoietin to increase the overall number of functional red blood cells of an organism, e.g., in renal anaemia. For example, during dialysis, red blood cells can be destroyed so that dialysis patients can become anaemic. Since the kidneys of these patients are often insufficient and non-functional, a proper erythropoietin supply is not guaranteed so that dialysis patients need rhuEpo therapy to provide a sufficient rhuEpo level, which then is able to continuously replace the destroyed red blood cells.

In the pharmaceutical compositions and uses of the invention the polypeptide of the invention may be coupled covalently or non-covalently to carriers, e.g., keyhole limpet hemocyanine and/or any effector system such as ricin or ^{99}Tc .

Figure 1: Biological activity of different concentrations of rhuEpo(WT) and rhuEpo(Gln24)

- A. The biologic activities of different concentrations of rhuEpo wt and rhuEpo(Gln24) were tested in a human bone marrow red colony assay. GM-CSF and medium samples were coanalyzed as controls. The number of red colonies was evaluated after two weeks of cultivation for each sample.

- B. The biological activities of different concentrations of rhuEpo wt and rhuEpo(Gln24) of the invention were tested in a human bone marrow red colony assay. Medium samples were coanalyzed as controls. The number of red colonies was evaluated after two weeks of cultivation for each sample.

The examples illustrate the invention.

Example 1: Construction of expression vector pHOEBE-40-7 (rhuEpo(Gln24))

For the expression of erythropoietin in CHO cells an expression system derived from the BPV-1 expression vector pCES was used¹¹. To optimize the translational start site according to Kozak¹², an ACC Triplet was inserted directly upstream of the first ATG codon of the Epo coding sequence by site directed mutagenesis. Then rhuEpo(wt)-coding sequences of vector pCES (BamHI - BglII fragment) were exchanged by the Epo sequences with the optimal transcriptional start site, resulting in expression vector pHOEBE-40-1. Alternatively, the sequence coding for mutein rhuEpo(Gln24), also including the Kozak sequence, was cloned into the expression vector, resulting in pHOEBE-40-7. In further experiments the pABE-40 expression vector system⁴ was used for the expression in CHO cells. The mutein sequences and the Kozak sequence were obtained by site directed mutagenesis¹³ and the sequences had been verified by sequence analysis. The BPV-1 expression system pHOEBE-40-contains DNA fragments of different origin. The different elements are

- a) a Sall - EcoRI fragment of 2317 bp derived from plasmid pJYM¹⁴, containing sequences of the bacterial plasmid pML-1, including the ampicillin resistance gene (Amp) and bacterial origin of replication (E.coli ORI);
- b) an EcoR I - BamH I fragment of 2801 bp, containing the mouse metallothionein 1 (MT-1) promoter and the 3' non-coding region of the murin MT-1 gene, in th

reverse orientation. Due to the cloning strategy, the two parts of the MT-1 gene, are separated by a short pBR322-derived sequence of 31 bp (Hind III - EcoR I) of vector pJYM¹⁴ ;

- c) a BamH I - Bgl II fragment of 766 bp, containing the rhuEpo(Gln24) c-DNA (E40-7)⁴, and the upstream optimized translation start site.
- d) a Bgl II - BamH I fragment of 242 bp containing the SV 40 late polyadenylation signal¹⁵ ;
- e) a BamH I - Sal I fragment of 7953 bp containing the total genomic sequence of the bovine papilloma virus BPV-1 (corresponding to the 100 % BPV-1 genome BamH I-fragment)¹⁶. This sequence contains a eukaryotic origin of replication, located in a fragment of about 100 bp (7900bp - 52bp of BPV-1 genome, adjacent to the Hpa I site at position 1). Furthermore, the sequence comprises open reading frames coding for BPV-1 - specific replication factors, transcription factors and transforming factors. It is suggested that these factors are able to modulate posttranslational modification processes.

Example 2: Transfection of CHO dhfr⁻ cells and detection of transient expression levels

CHO dhfr⁻ cells were grown in Dulbecco's modified Eagle's medium (DMEM) containing 10 % fetal calf serum (FCS). Transfections of CHO dhfr⁻ cells were carried out using the method according to Graham and Van der Eb¹⁷. Transient expression and secretion was analyzed 24 , 48, and 72 hours after transfection of 10 µg/ml of rhuEpo expression vector DNA (pABE-40-7)⁴. In each transient expression experiment, vector p4EGD¹⁸ carrying the coding sequence of a truncated human IgG₁ Fc (rhu IgG₁ Fc) under the control of the SV40 promoter^{19,20} was cotransfected, and the expression rates of both rhuEpo and rhu IgG₁ Fc were determined using specific ELISAs^{21,10}, respectively. The relative secretions of the rhuEpo muteins were standardized on the secretion rates of rhuEpo(wt) and rhu IgG₁ Fc (see Table I).

TABLE I: Transient Secretion of CHO-Derived rhuEpo Muteins

	1	2		3	4
Time (hours p.t.)	rhuEpo Mutein	Absolute Secretion (ng/ml)		Secretion (%)	Secretion (%)
		rhuEpo	Fc	Relative to Fc	Relative to rhuEpo(wt)
24	rhuEpo(wt)	2.9	4.9	59.2	100.0
	rhuEpo(Gln24)	23.0	13.7	167.9	283.6
48	rhuEpo(wt)	7.2	14.0	51.4	100.0
	rhuEpo(Gln24)	53.0	21.0	252.4	491.0
72	rhuEpo(wt)	9.7	23.0	42.2	100.0
	rhuEpo(Gln24)	74.0	31.0	238.7	565.6

Transient transfectants were tested for expression of rhuEpo(wt) and rhuEpo(Gln24) relative to IgG₁ Fc fusion protein as a secreted reference protein post transfection (p.t.)

at 24 h, 48 h and 72 h, respectively. The average values of triplicate determinations (n=3) of the absolute secretion levels from each transfection set were averaged. The variations of the triplicate values were below 5 % for either rhuEpo/rhuEpo mutein secretion or rhuEpoRFc secretion (column 2). From these data the rhuEpo expression levels relative to IgG₁Fc (Fc, 100%) expression were calculated on a percent basis for each experiment (column 3). From these values, the rhuEpo(Gln24) secretion levels were estimated relative to rhuEpo(wt) on a percent basis for each experiment (column 4).

Example 3: Detection of transient rhuEpo(wt), rhuEpo(Gln24) mutein and rhuIgG₁ Fc expression levels

Supernatants of transiently or stably transfected cultures were tested for rhuEpo(wt) or rhuEpo mutein content by a rhuEpo-specific ELISA^{21,10}. Based on a polyclonal rabbit antiserum, this assay was carried out as follows: microtitration plates were coated overnight with 500 ng per ml of a rabbit anti-rhuEpo immunoglobulin fraction. Then the plates were washed three times with PBS containing 0.01 % of Tween20 and air-dried at 37 °C. Before use, the plates were saturated with PBS containing 0.05 % (w/v) of bovine serum albumin (BSA). Then the plates were washed three times with PBS-Tween20 and incubated with the supernatant samples for 30 minutes. After washing for three times with PBS containing 0.05 % (w/v) of BSA, bound rhuEpo or rhuEpo muteins were detected by peroxidase-labelled rabbit anti-rhuEpo immunoglobulin fraction (Ig-POD). After 30 min of incubation, the microtitration plates were washed and the remaining peroxidase activity, corresponding to captured rhuEpo, was developed, using tetramethylbenzidine (TMB) as a substrate. Then the reaction was stopped by addition of H₂SO₄, and the plates were measured in a Behring ELISA Processor II (Behringwerke AG, Marburg, FRG).

Supernatants of transiently transfected cell cultures were screened for the secretion of IgG₁Fc, to determine the relative secretion of the different rhuEpo muteins. For this, microtitration plates were coated with goat anti-human Fc polyclonal immunoglobulin fraction. After washing and saturation (see above), the plates were incubated with the supernatants for 1 hour. Then the captured IgG₁Fc molecules were detected by goat anti-human Fc antibodies, labelled with peroxidase (POD) as described above.

Example 4: Production of stable rhuEpo(Gln24) mutein expressing cell clones

Cell clones secreting rhuEpo(Gln24) mutein were obtained by cotransfection of the vector pSV2 dhfr expressing the dihydrofolatereductase gene, providing resistance against methotrexate¹⁸, together with expression vectors pABE-40-1, pABE-40-7, pHOEBE-40-1, or pHOEBE-40-7, respectively. After transfection the cultures were selected in the presence of methotrexate. After a period of two to three weeks, cell colonies grew out and single cell clones were cloned by use of cloning cylinders or according to the limiting dilution method. The production of rhuEpo(Gln24) was analyzed as described in Example 2.

Example 5: Upscaling of production cell clones

Cell clones suitable for production were further cultivated and finally expanded to roller bottle cultures. To produce erythropoietins, cells from the respective seed lots were expanded in roller bottles to confluence. Then the growth medium was replaced by serum-free DMEM, which was harvested at the end of the production phase.

Example 6: Purification of rhuEpo(Gln24)

Purified rhuEpo and glycomutins were obtained by affinity chromatography using monoclonal antibody 146/0056¹⁰. This antibody was covalently coupled to sepharose CL4B (Pharmacia; Uppsala, Sweden) according to Fibi¹⁰. After elution at pH 2.5 into 1 ml of 1M Tris-HCl, pH 9.5, the samples were dialyzed against PBS pH 7.0. The protein concentration was calculated from the O.D. 280 nm, and the purity of the preparation was controlled visually after separation in a polyacrylamide gel and subsequent silver staining (Phast System, Pharmacia).

Example 7: SDS-Page and silver staining

SDS-PAGE and Western blotting procedures were carried out as described recently¹⁰, using the Phast System, except that rainbow marker proteins and a gold-labelled-antibody procedure (both from Amersham-Buchler, Braunschweig, FRG) were used.

Example 8: Human erythroid precursor colony assay

Suspensions of human bone marrow cells were prepared from bone marrow specimens in phosphate buffered saline. Gradient-purified interphase cells of the suspensions were harvested after centrifugation at 2200 rotations per minute for 25 min at 12 °C. The cells were washed three times, resuspended in MEM-alpha medium with supplements and certomycin as an antibiotic. The cell number was adjusted to 1.1×10^6 / ml, and the cells were mixed with 4 ml rhuEpo Medium (19,1 ml FCS / 15,2 ml Transferrin/ BSA/ FeCl₃ / 13,7 ml MEM-alpha medium), and 0,7 ml Agar (about 70 °C). 200 µl of the agar cell suspension were added per well to 24 well tissue culture plates, containing dilutions of the test samples. The cultures were mixed with the test samples and incubated for 14 days at 37°C in an atmosphere containing 7 % CO₂ and 10 % O₂.

Example 9: Glycoanalysis

The liberation by PNGase F of the N-glycans of rhuEpo was performed as described by Nimtz.⁷ The liberated N-glycan pools were measured by high-pH anion-exchange chromatography with pulsed amperometric detection (HPAE-PAD), using the set-up and the optimized standard gradient "S" for sialylated glycans previously described²². The hypothetical N-glycan charge Z was determined as described by Hermentin^{23,24}. In brief, the hypothetical N-glycan charge of the rhuEpo samples was gained by

- i) liberating the N-glycan-pool of the glycoprotein via PNGase F
- ii) measuring the N-glycan pool via HPAE-PAD
- iii) calculating the percentage of the areas (A) of the groups of peaks, separated by charge,
- iv) multiplying the area% of the peak-groups in the neutral (asialo-, as), monosialo- (MS), disialo- (DiS), trisialo- (TriS), tetrasialo- (TetraS) and pentasialo (PentaS) region by zero (asialo), 1 (MS), 2 (DiS), 3 (TriS), 4 (TetraS), and 5 (Sulfated), respectively, and
- v) summarizing the respective products.

Thus, Z was defined as the sum of the products of the respective areas (A) in the asialo (as), monosialo (MS), disialo (DiS), trisialo (TriS), tetrasialo (TetraS) and sulfated region, each calculated as the percentage of the total peak area set equal to 100%, and each multiplied by the corresponding charge:

$$Z = A(\text{as}) * 0 + A(\text{MS}) * 1 + A(\text{DiS}) * 2 + A(\text{TriS}) * 3 + A(\text{TetraS}) * 4 + A(\text{Penta S or Sulfated}) * 5$$

- vi) dividing Z through the number of glycosylation sites to create Z*. Z* gives an estimate of the number of charges per N-glycosylation site and, thus, of the grade of chargement of a glycosylation site.

The results of the glycoanalysis of rhuEpo so far commercially available and the rhuEpo of the invention (column C4 and C5) are compared in Table II.

Table II: Comparison of the glycosylation profiles of different recombinant human erythropoietins

	Organon rhuEpo (CHO) pABE-40-like		Amgen* rhuEpo (CHO) pABE-40-like		Boehringer rhuEpo (CHO) pABE-40-like		Behring-Mutein rhuEpo (Gln24) (CHO) pABE-40-7	
							C3	
area of intergration (peak group)	peak group area (%)	charge number share	peak group area (%)	charge number share	peak group area (%) O950579 K.D07	charge number share	peak group area (%) O950576 K.D04	charge number share
asialo							3,1	0
monosialo							6,1	6,0
disialo	15	30,0	6,4	12,8	5,4	10,7	11,3	23,0
trisialo	32	96,0	20,7	62,1	25,5	76,5	23,7	71,0
tetrasialo	40	160,0	72,9	291,6	69,2	276,6	55,9	223,0
sulfated			n.d.		n.d.		0,0	0
N-glycan charge number Z (total)	(a)	286	(b)	367		364		323
Z*		95,3		122,3		121,3		162,5

	Behring-Mutein rhuEpo (Gln24) (CHO) pHOEBE-40-7				Merckle rhuEpo (BHK) pABE-40-like			
	C4		C5				Nimtz	
area of intergration (peak group)	peak group area (%) O950576 K.D04	charge number share	peak group area (%) O950546 K.D20	charge number share	peak group area (%)	charge number share	peak group area (%)	charge number share
asialo								
monosialo	2,8	2,8	1,8	1,8	4,7	4,7	n.d.	
disialo	3,2	6,3	8,6	17,3	14,5	29,0	21,1	42,2
trisialo	13,8	41,4	20,2	60,6	33,9	101,7	35,0	105,0
tetrasialo	64,0	256,0	60,6	242,4	46,8	187,2	40,9	163,6
sulfated	16,2	81,2	8,7	43,5				
N-glycan charge number Z (total)		388		366	(c)	323	(d)	311
Z*		194		183		107		103

- a) calculated from Hokke²⁵
 - b) calculated from Watson²⁶
 - c) average of 4 different batches; 1 HPAE-PAD run, each
 - d) calculated from Nimtz⁷
- Z* Total N-glycan charge number Z / number of N-glycosylation sites.
- n.d. not determined

The carbohydrate analysis of rhuEpo, originally described by Sasaki et al.¹ and Takeuchi²⁷ for rhuEpo (CHO) and by Tsuda²⁸ for rhuEpo (BHK), has recently been extended by studies of Hokke²⁵, Watson²⁶ (for CHO-rhuEpo), and Nimtz⁷ (for BHK-rhuEpo). According to a new method, the hypothetical N-glycan charge Z can be determined as described by Hermentin^{23,24}. It has been demonstrated that this parameter gives an excellent estimate of the amount of undersialylated N-glycans to properly sialylated N-glycans. As the glycans of rhuEpo are known to consist of mainly tetraantennary structures with 0-3 LacNAc repeats, Z should amount to a hypothetical N-glycan charge number between 300 and 400. Indeed, the N-glycan charge of CHO-rhuEpo (Boehringer Mannheim) was determined to $Z = 364 \pm 2$ (CV = 0.6%) (n = 6; three different experiments with 2 HPAE-PAD runs, each), and the N-glycan charge of BHK-rhuEpo (Merckle) was determined to $Z = 323 \pm 2$ (CV = 0.7%) (n = 4; four different lots; 4 different experiments; 1 HPAE-PAD run, each); see Table II²⁴. Thus, the smaller Z value of the BHK-rhuEpo from Merckle clearly reflected the greater share of undersialylated N-glycans: 34% of the N-glycans were missing one and 12% of the N-glycans were missing two terminal sialic acid residues; the structures consisted of 40.9 % tetrasialylated, 35.0 % trisialylated and 21.1 % disialylated structures (Nimtz.⁷). These data from the literature allowed to calculate the N-glycan charge to $Z = 311$, which is in good agreement (deviation <4%) with the N-glycan charge determined according to Eq. 1, supra, i. e., $Z = 323$, using the same rhuEpo (BHK) from Merckle (see Table II). In the CHO-rhuEpo from Amgen the major (>95%) di-, tri- and tetra-antennary structures were fully sialylated²⁶. Their separation according to charge of the

PNGase F-released N-glycans (using a Glycopak DEAE column) allowed to calculate the N-glycan charge to $Z = 367$, which is in excellent agreement with the glycan charge of the rhuEpo (CHO) from Boehringer Mannheim, used in this study ($Z = 364$, see Table II). In contrast, the study of Hokke et al.²⁵, investigating CHO-rhuEpo from Organon Teknika, showed that 18-20% of the N-glycans were missing one, and 3% of the N-glycans were missing two sialic acid residues. Their structural analysis enabled to calculate the N-glycan charge to $Z = 286$, which is significantly smaller than the CHO-rhuEpo from Amgen ($Z = 367$) or Boehringer ($Z = 364$); see Table II)²⁴.

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Claims

1. A polypeptide having part or all of the primary structural conformation of erythropoietin possessing the biological property of causing bone marrow cells to increase production of reticulocytes and red blood cells and to increase hemoglobin synthesis or iron uptake, said polypeptide being the product of eukaryotic expression of an exogenous DNA sequence and having the following physiochemical properties:
 - (i) the amino acid sequence comprises the amino acid sequence given in SEQ ID No. 1 or any fragment or derivative thereof by way of amino acid deletion, substitution, insertion, addition and/or replacement of the amino acid sequence given in SEQ ID No. 1, wherein at least one of the consensus N-linked glycosylation sites is modified to other than a consensus N-linked glycosylation site;
 - (ii) it is glycosylated;
 - (iii) greater 5% of the N-glycan structures are sulfated; and
 - (iv) the ratio Z^* of the total N-glycan charge Z to the number of N-glycosylation sites is greater than 170.
2. The polypeptide of claim 1, wherein at least one of the consensus N-linked glycosylation sites is deleted.
3. The polypeptide of claim 1 or 2, wherein at least one of the consensus N-linked glycosylation sites is replaced with (a) different amino acid(s).
4. The polypeptide of any one of claims 1 to 3, wherein at least one of the consensus N-linked glycosylation sites is added.
5. The polypeptide of any one of claims 1 to 4, wherein the glycosylation site at amino acid Asn24 of the amino acid sequence given in SEQ ID No. 1 is deleted.

6. The polypeptide of any one of claims 1 to 5, wherein the amino acid Asn at position 24 of the amino acid sequence shown in SEQ ID No. 1 has been replaced with the amino acid Gln.
7. The polypeptide of claim 6 having the amino acid sequence as shown in SEQ ID No. 2.
8. The polypeptide of any one of claims 1 to 7, wherein Z^* is greater than 180.
9. The polypeptide of any one of claims 1 to 8, wherein Z^* is greater than 190.
10. A DNA sequence encoding the amino acid sequence of a polypeptide of any one of claims 1 to 9 operatively linked to regulatory elements of Bovine Papillomavirus 1 (BPV-1) which allow for the expression of said DNA in a eukaryotic host cell.
11. The DNA sequence of claim 10, further comprising regulatory elements of the metallothioneine1 (MT-1) gene.
12. An expression vector comprising the DNA sequence of claim 10 or 11.
13. The expression vector of claim 12, which is BPV-1 vector pHOEBE-40-7.
14. A eukaryotic host cell comprising the DNA sequence of claim 10 or 11 or the vector of claim 12 or 13.
15. The host cell of claim 14, which is a CHO cell.

16. A method for the production of a polypeptide as defined in any one of claims 1 to 9 having part or all of the primary structural conformation of erythropoietin possessing the biological property of causing bone marrow cells to increase production of reticulocytes and red blood cells and to increase hemoglobin synthesis or iron uptake, said method comprising culturing a host cell of claim 14 or 15, and optionally isolating said polypeptide from the culture.
17. A polypeptide obtainable by the method of claim 16.
18. The polypeptide of any one of claims 1-9 or 17, characterized in that it displays the following feature(s):
- (a) increased half life in vivo; and/or
 - (b) increased bioactivity compared to the same polypeptide having a Z* less than 170.
19. A pharmaceutical composition comprising a polypeptide of any one of claims 1 to 9 or of claim 17 or 18, and optionally a pharmaceutically acceptable carrier.
20. A pharmaceutical composition comprising a DNA sequence of claim 10 or 11 and/or a vector of claim 12 or 13, and optionally a pharmaceutically acceptable carrier.
21. A diagnostic composition comprising a polypeptide of any one of claims 1 to 9 or of claim 17 or 18, and optionally suitable means for detection.
22. Use of the polypeptide of any one of claims 1 to 9 or of claim 17 or 18 for the preparation of a pharmaceutical composition for treating all kinds of anemia caused by a lack of erythropoietin.

23. Use of the DNA sequence of claim 10 or of the vector of claim 12 or 13 for the preparation of a pharmaceutical composition for treating all kinds of anemia caused by a lack of erythropoietin.

Figure 1A

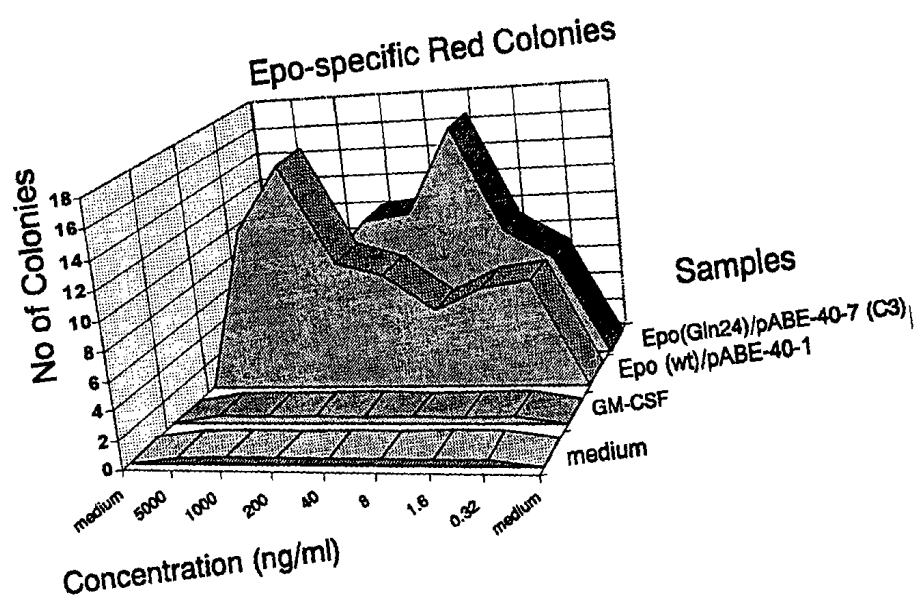
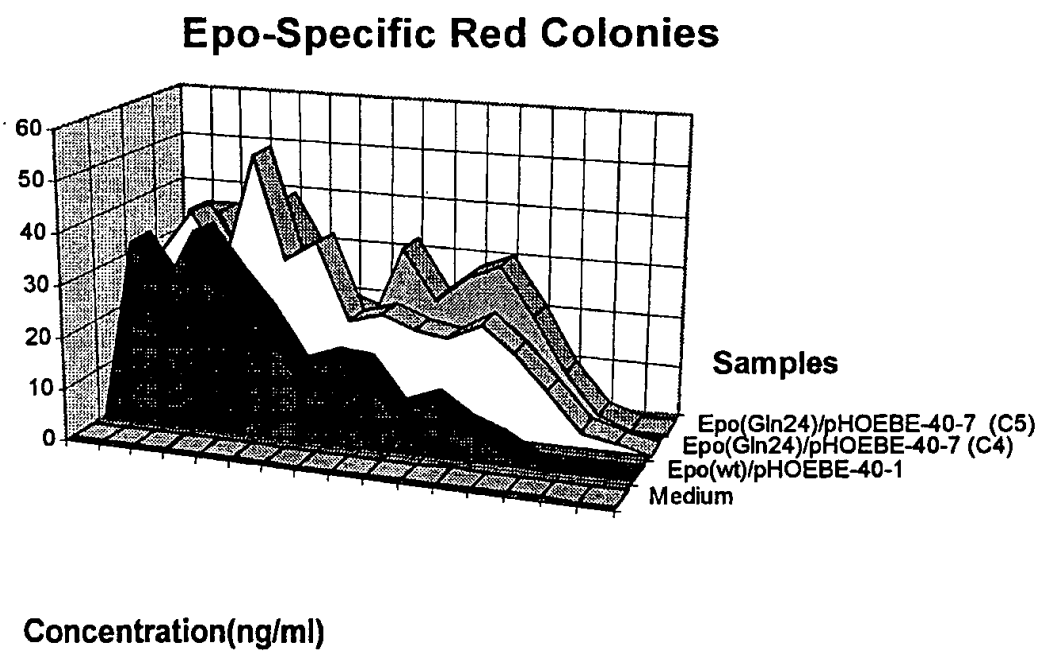


Figure 1B



SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

(A) NAME: Hoechst Marion Roussel Deutschland GmbH
(B) STREET: -
(C) CITY: Frankfurt
(D) STATE: -
(E) COUNTRY: Germany
(F) POSTAL CODE (ZIP): 65926
(G) TELEPHONE: 069-305-3005
(H) TELEFAX: 069-35-7175
(I) TELEX: -

(ii) TITLE OF INVENTION: Recombinant Human Erythropoietin With
Advantageous Glycosylation Profile

(iii) NUMBER OF SEQUENCES: 2

(iv) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 166 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

Ala	Pro	Pro	Arg	Leu	Ile	Cys	Asp	Ser	Arg	Val	Leu	Glu	Arg	Tyr	Leu
1				5					10					15	
Leu	Glu	Ala	Lys	Glu	Ala	Glu	Asn	Ile	Thr	Thr	Gly	Cys	Ala	Glu	His
			20					25					30		
Cys	Ser	Leu	Asn	Glu	Asn	Ile	Thr	Val	Pro	Asp	Thr	Lys	Val	Asn	Phe
			35				40					45			

Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln Ala Val Glu Val Trp
 50 55 60
 Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu Arg Gly Gln Ala Leu
 65 70 75 80
 Leu Val Asn Ser Ser Gln Pro Trp Glu Pro Leu Gln Leu His Val Asp
 85 90 95
 Lys Ala Val Ser Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu
 100 105 110
 Arg Ala Gln Lys Glu Ala Ile Ser Pro Pro Asp Ala Ala Ser Ala Ala
 115 120 125
 Pro Leu Arg Thr Ile Thr Ala Asp Thr Phe Arg Lys Leu Asp Arg Val
 130 135 140
 Tyr Ile His Pro Phe Arg Gly Lys Leu Lys Leu Tyr Thr Gly Glu Ala
 145 150 155 160
 Cys Arg Thr Gly Asp Arg
 165

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 166 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Ala Pro Pro Arg Leu Ile Cys Asp Ser Arg Val Leu Glu Arg Tyr Leu
 1 5 10 15
 Leu Glu Ala Lys Glu Ala Glu Gln Ile Thr Thr Gly Cys Ala Glu His
 20 25 30
 Cys Ser Leu Asn Glu Asn Ile Thr Val Pro Asp Thr Lys Val Asn Phe
 35 40 45
 Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln Ala Val Glu Val Trp

50	55	60
Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu Arg Gly Gln Ala Leu		
65	70	75 80
Leu Val Asn Ser Ser Gln Pro Trp Glu Pro Leu Gln Leu His Val Asp		
	85	90 95
Lys Ala Val Ser Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu		
	100	105 110
Arg Ala Gln Lys Glu Ala Ile Ser Pro Pro Asp Ala Ala Ser Ala Ala		
	115	120 125
Pro Leu Arg Thr Ile Thr Ala Asp Thr Phe Arg Lys Leu Asp Arg Val		
	130	135 140
Tyr Ile His Pro Phe Arg Gly Lys Leu Lys Leu Tyr Thr Gly Glu Ala		
145	150	155 160
Cys Arg Thr Gly Asp Arg		
	165	

INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 98/05399

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/12 C07K14/505 A61K38/18 A61K31/70 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K A61K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the International search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	FIBI M.R. ET AL.: "N- and O-glycosylation mutants of recombinant human erythropoietin secreted from BHK-21 cells." BLOOD, vol. 85, no. 5, 1995, pages 1229-1236, XP002053700 cited in the application * the whole document, especially page 1235, last paragraph of discussion *	1-23
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☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- "&" document member of the same patent family

Date of the actual completion of the international search

7 January 1999

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

Int. l. Application No

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Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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International Application No

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Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	FIBI M.R. ET AL.: "Inactivation of recombinant plasmid DNA from a human erythropoietin-producing mouse cell line grown on a large scale." APPL. MICROBIOL. BIOTECHNOL., vol. 35, 1991, pages 622-630, XP002053705 cited in the application see figure 1 see page 623, left-hand column, last paragraph - right-hand column, paragraph 2 -----	10-14, 16,20,23
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